

Synthesis of Epiantillatoxin, a Stereoisomer of the Potent Ichthyotoxin from *Lyngbya majuscula*

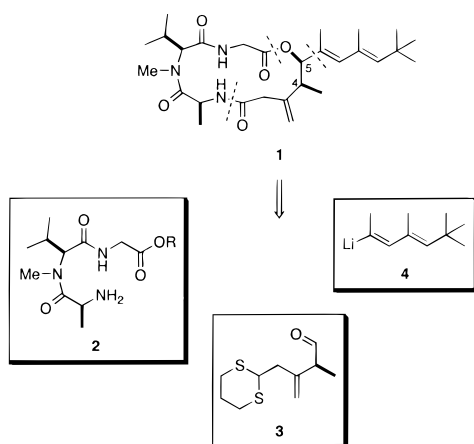
James D. White,* Roger Hanselmann, and Duncan J. Wardrop

Department of Chemistry, Oregon State University,
Corvallis, Oregon 97331-4003

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Cyanobacteria (blue-green algae) have proven to be a rich source of novel marine metabolites.¹ A collection of *Lyngbya majuscula* near Curaçao furnished, in addition to the novel tubulin-binding agent curacin A,² a small amount of a powerful ichthyotoxic substance named antillatoxin (LD₅₀ = 0.005 mg/mL). Structure **1**, including the (4*S*,5*R*) configuration, was attributed to antillatoxin by Gerwick et al. on the basis of ¹H NMR spectroscopy, analysis of its CD spectrum, and molecular modeling.³ Herein, we report a synthesis of **1** which establishes that this assignment to antillatoxin is incorrect.⁴

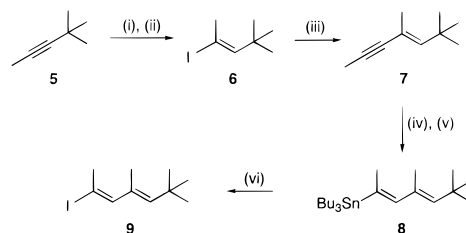
Our synthetic approach follows a convergent strategy which assembles **1** from the tripeptide **2**, an aldehyde **3**, and 1,3,5,5-tetramethylhexa-1,3-dienyllithium (**4**). After the C5–C6 bond is



established, the coupled product is connected to the carboxyl terminus of **2** prior to final lactamization.⁵

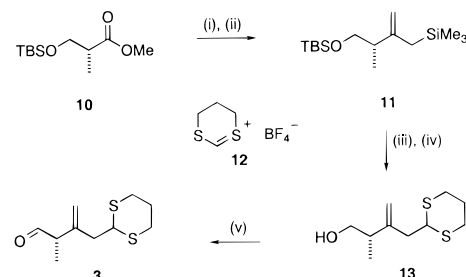
Synthesis of the diene side chain **4** of antillatoxin began from 4,4-dimethylpent-2-yne (**5**), which was subjected to hydrozirconation with Schwartz' reagent⁶ and the resultant vinylzirconocene species was reacted with *N*-iodosuccinimide to yield iodoalkene **6**.⁷ The latter was coupled with propynylmagnesium bromide in the presence of a palladium(0) catalyst⁸ to give **7**. Stannylcupration of enyne **7** at low temperature, followed by methanolysis, led to **8**, accompanied by ca. 10% of the inseparable regioisomeric dienylstannane.⁹ Iodination of the mixture furnished pure iododiene **9** after flash chromatography (see Scheme 1).

Scheme 1^a



^a (i) Cp₂ZrClH, THF; (ii) *N*-iodosuccinimide; (iii) MeC≡CMgBr, Pd(PPh₃)₄ (2.5 mol %), THF, 68% from **5**; (iv) Bu₃Sn(Bu)CuCNLi₂, THF, -50 °C; (v) MeOH, -50 °C → -10 °C, overnight; (vi) I₂, Et₂O, 0 °C, 52% from **7**.

Scheme 2^a



^a (i) Me₃SiCH₂MgCl, CeCl₃, THF; (ii) SiO₂, CH₂Cl₂, 99%; (iii) **12**, CH₃NO₂, 0 °C; (iv) BF₃·OEt₂, 0 °C, 70%; (v) IBX (1 M in DMSO), THF, 70%.

The C1–C5 subunit **3** of **1** was constructed from methyl (*R*)-3-hydroxy-2-methylpropionate via a sequence which conveniently differentiated the two aldehyde termini of this segment. Addition of excess (trimethylsilyl)methylmagnesium chloride to **10** in the presence of cerium trichloride gave a tertiary carbinol which, upon exposure to silica, underwent Peterson elimination to furnish allylsilane **11**.¹⁰ Electrophilic substitution of **11** with 1,3-dithienium fluoroborate (**12**)¹¹ in nitromethane produced a mixture of alcohol **13** and its TBS ether (ca. 1:1); the mixture was converted directly to **13** upon treatment with boron trifluoride etherate (see Scheme 2). Oxidation of **13** with *o*-iodoxybenzoic acid (IBX)¹² furnished the unstable aldehyde **3** which was reacted immediately with the lithiated diene **4**, obtained from **9** by treatment with *tert*-butyllithium at low temperature, to yield an 8:1 mixture of two stereoisomeric alcohols. After chromatographic purification, the major alcohol **14** was acetylated, and the acetate **15** was converted to carboxylic acid **16** by mild cleavage of the dithiane¹³ followed by oxidation of the resultant aldehyde with buffered sodium chlorite. The relative configuration of **16** was established by its transformation to δ -lactone **17** in which the *cis* relationship of hydrogens at C4 and C5 was confirmed by their coupling constant of 3.0 Hz and a NOE of 7%. This result established the configuration of the major alcohol from the reaction of **3** with **9** as (5*R*), corresponding to the Felkin mode of addition (see Scheme 3).

The tripeptide unit of antillatoxin was prepared from Cbz-protected *N*-methylvaline (**18**)¹⁴ by bromotris(dimethylamino)-phosphonium hexafluorophosphate(BroP)-mediated coupling with methyl glycinate to give dipeptide **19**. The Cbz group of **19** was

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(5) An unsuccessful approach to **1** along different lines has been reported (Loh, T.-P.; Cao, G.-Q.; Pei, J. *Tetrahedron Lett.* **1998**, *39*, 1453, 1457).

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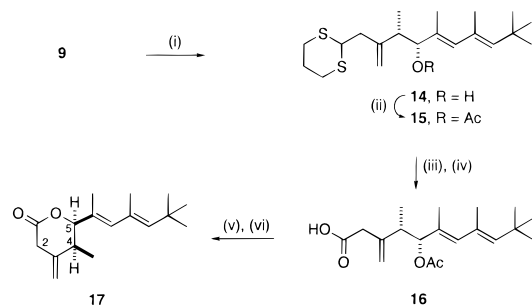
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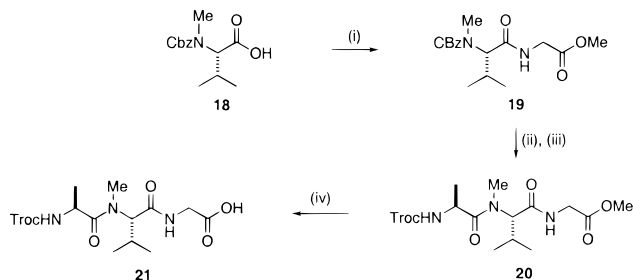
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Scheme 3^a

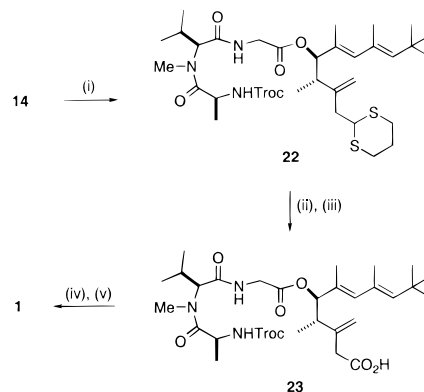
^a (i) *t*-BuLi, THF, $-78\text{ }^{\circ}\text{C}$, 20 min, then **3**, 60%; (ii) Ac_2O , py, 20 h, 100%; (iii) MeI, CaCO_3 , $\text{MeCN}-\text{H}_2\text{O}$, 16 h; (iv) NaClO_2 , NaH_2PO_4 , $\text{MeCH}=\text{CMe}_2$, *t*-BuOH- H_2O , 91%; (v) K_2CO_3 , MeOH; (vi) CSA, CH_2Cl_2 , 43%.

Scheme 4^a

^a (i) Gly-OMe, BroP, (*i*-Pr)₂NEt, CH_2Cl_2 , 93%; (ii) H_2 , Pd/C, MeOH; (iii) Troc-ala OH, HATU, (*i*-Pr)₂NEt, CH_2Cl_2 , 74%; (iv) LiOH, THF, 100%.

removed by hydrogenolysis, and the amine was condensed, using (*O*-7-azabenzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HATU) as the coupling agent, with 1,1,1-trichloroethoxycarbonyl (Troc)-protected alanine.¹⁵ The resultant tripeptide **20** was saponified to furnish carboxylic acid **21** (see Scheme 4). Esterification of **14** with **21** was carried out in the presence of 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide (EDC) and afforded **22** in good yield. After removal of the dithiane, the derived aldehyde was oxidized to carboxylic acid **23**. The Troc-protecting group was cleaved, and final macrolactamization of the liberated amino acid using HATU produced **1** (see Scheme 5). The latter was not identical in spectral properties, optical rotation, or chromatographic behavior with a sample of natural antillatoxin, and it is therefore concluded that the structure assigned³ to antillatoxin must be revised.

Although most features of the ¹H and ¹³C NMR spectra of our synthetic **1** are similar to those of natural antillatoxin and are

Scheme 5^a

^a (i) **21**, EDC, DMAP, CH_2Cl_2 ; (ii) MeI, CaCO_3 , $\text{MeCN}-\text{H}_2\text{O}$; (iii) NaClO_2 , NaH_2PO_4 , $\text{MeCH}=\text{CMe}_2$, *t*-BuOH- H_2O , 75% from **14**; (iv) Zn, KH_2PO_4 , THF; (v) HATU, (*i*-Pr)₂NEt, DMF, 60% from **23**.

consistent with a stereoisomeric relationship, conspicuous differences are the chemical shift of H4 (δ 2.71 in **1**, δ 2.17 in antillatoxin) and the coupling constant between H4 and H5. The observed value of this coupling in **1** ($J \approx 0$ Hz) is indicative of a *cis* orientation of hydrogens, whereas the reported value ($J = 11$ Hz) for antillatoxin clearly is not. *Cis* configuration at C4,C5 in the natural product was assigned by Gerwick on the basis of a NOE between protons attached to these carbons,³ but reexamination of the NOE experiment has found that this result is spurious.¹⁶ NMR evidence therefore points strongly toward a 4,5-*trans* configuration for antillatoxin. At this time, we are unable to distinguish between the (4*R*,5*R*) and (4*S*,5*S*) stereochemical assignments for the natural product.

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Supporting Information Available: Experimental procedures, characterization data, and NMR spectra (¹H and ¹³C) for synthetic intermediates (PDF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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